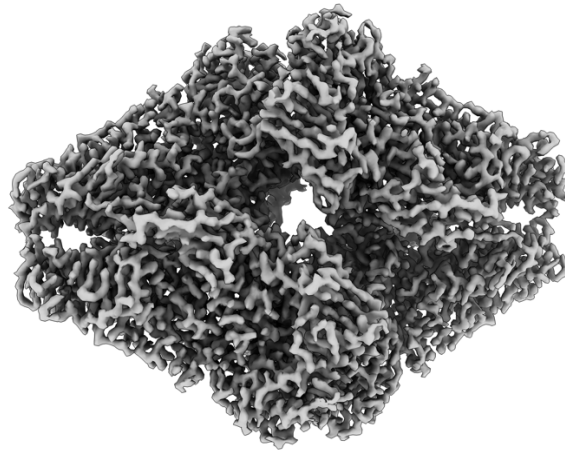


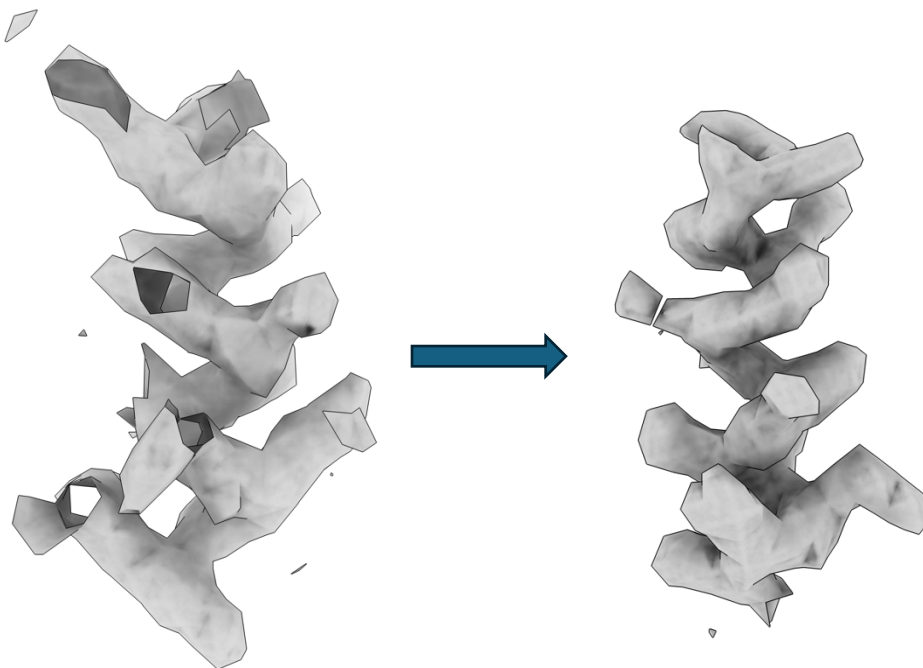
Cryo-EM Data Processing of β -Galactosidase in cryoSPARC 4.4 Tutorial Part II

- 1- Download the [sharp map](#) of b-gal you got during the 3D refinement of cryosparc and open it with chimeraX.
- 2- Create a working directory somewhere where you will put the map and models.

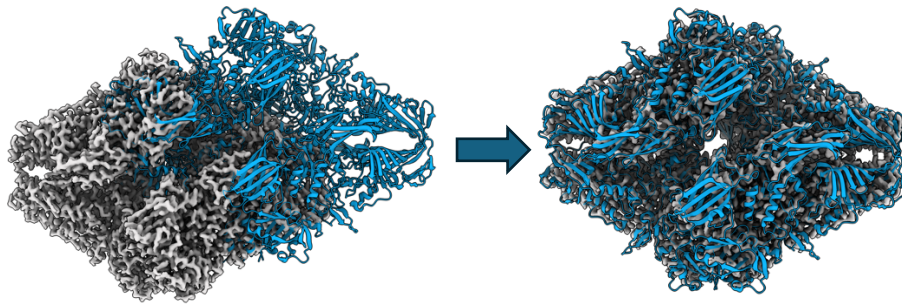


The first step is to check the quality of the map and its orientation. If the map does not appear satisfactory at first glance, there is no need to proceed further; you should continue with the data processing. If you observe secondary features, it's a positive sign and aligns with what we are seeking. Some of you may have an incorrectly oriented map (CryoSPARC does not consider the left-hand orientation as incorrect). If this occurs, you will need to manually flip the orientation.

The best way to determine this is by identifying a well-resolved alpha-helix: if you find one, use the following command in the script tab to flip the orientation: `vop flip #your_map_ID`

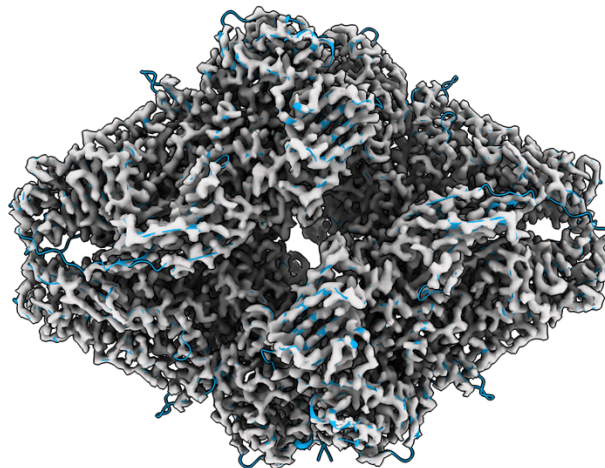


- 3- Download from the PDB a structure of b-gal (coming from ecoli | **P00722**), I used 8BKG and open the model with chimeraX. The first step is to align the PDB (manually, and roughly) the model with the map.



- 4- Once you reach this, you can tell chimeraX to perform a map fitting using the command:
`fitmap #model_ID inMap #map_ID`

It comprises a rigid body optimized for aligning an atomic model with a map. You may need to adjust and realign several times to achieve the perfect fit. While there are more advanced software options for this step, the visual method is typically preferred for its speed and ease of identifying any issues.



- 5- You know must save the model (which contains new coordinates after fitting into the map), and the flipped map you generated (if you have one). Using the command:
`save /Users/your_chosen_path/fitted-bgal.pdb #model_of_bgal`
`save /Users/your_chosen_path/z-flip_map.mrc #map_of_bgal`

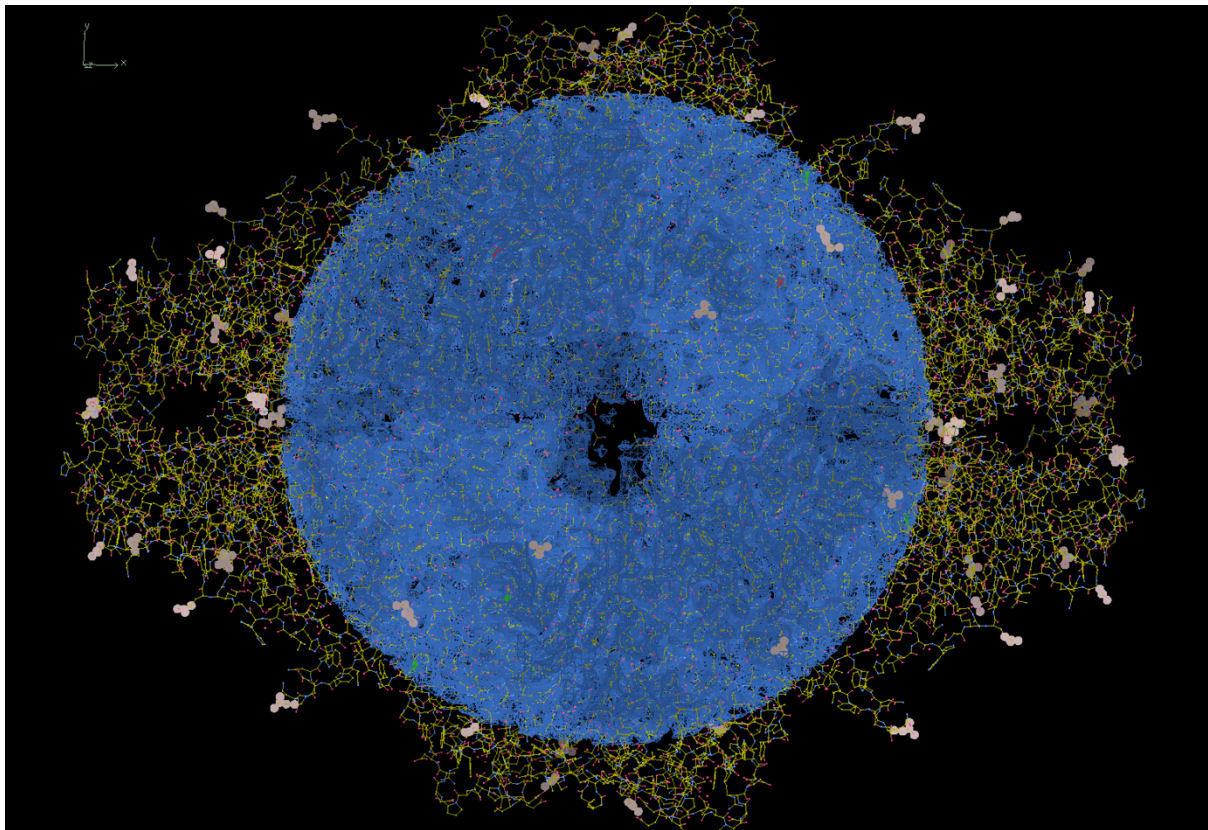
Please pay attention to the extension file: .pdb for the model and .mrc for the EM map.

- 6- We can look a bit closer into the map and the quality of fitting. Of course that will depend on your final map quality. For this, we will use Coot (same as what you did during the X-ray training).

Open coot

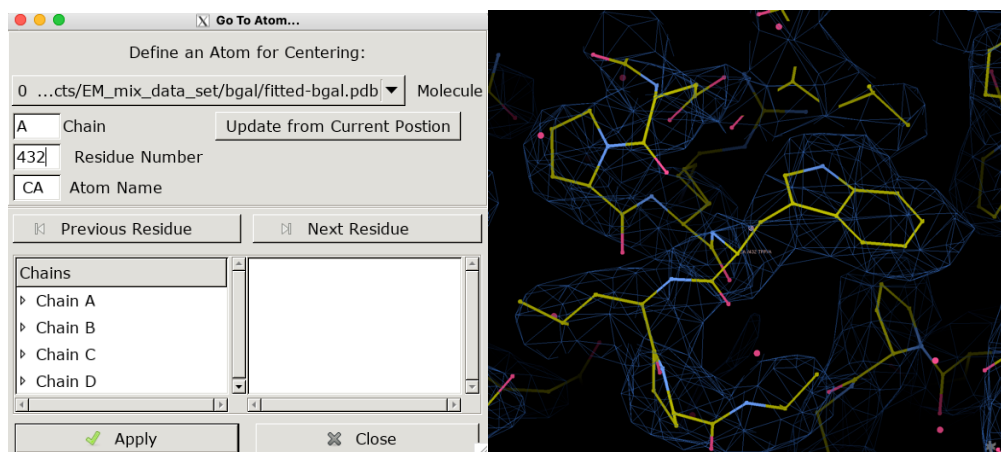
Select the model go to File > open coordinates > fitted-bgal.pdb

Select the map go to File > open map > z-flip_map.mrc



The cryoEM map is usually very large, and most laptops cannot handle the cache demanding to show everything. To avoid slow motion and crash you can quickly go to [Edit > Map parameters > Map Radius EM: 15A](#) and press [Apply](#).

Let's have a look around a nicely resolved (in most models) alpha-helix: 432-447. Go to atom a seek for this one and have a look around. Usually in cryoEM visual inspection (already from ChimeraX) is important.



If your final resolution is good enough, you should be able to see the TRP432 fitted into the cryoEM density. You can play with the level to see more or less the contrast.

If the fitting is not correct, you might want to go back to the previous steps and redo it again.

7- Once you are happy with the fitting, we will perform a Real Space Refinement of the model into the cryoEM map.

For this, you will open [Phenix](#), create a new project (where you saved your previous data) and select the job [real-space refinement](#).

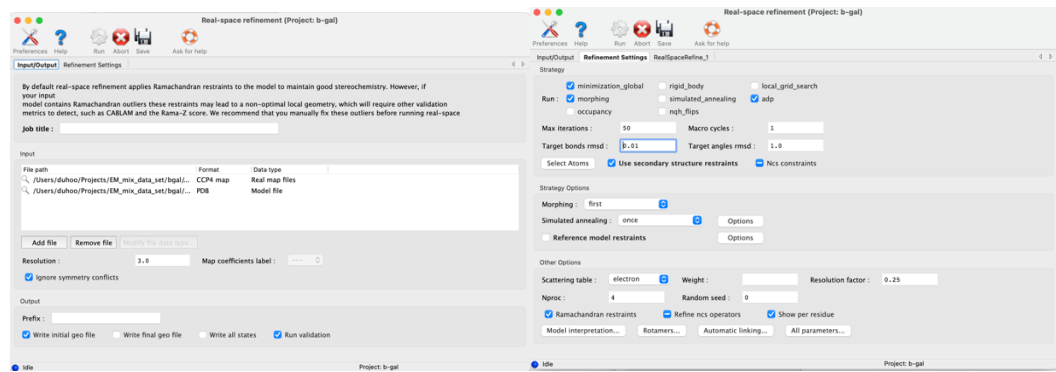
Select the EM_map and the PDB model.

Select the resolution you reached (and ignore symmetry conflicts).

Go to the Refinement Settings tab and select the same settings than the images.

We do not go into much detail here for a matter of time.

Press Run



- 8- Once you are done with the RSR, you can look at the validation report and start fixing the problem until your model matches the obtained data. We will not do this today.

Input/Output

Refinement Settings

RealSpaceRefine_1

Run status

Results

Validation

Summary

MolProbity

Model vs. Data

Data


Files

Model:

fitted-bgal_real_space_refined_001.pdb

Map:

cryosparc_P106_j177_007_volume_map_sharp_elflip.mrc



Open In Coot

Export Table 1

Model

Composition (#)

Chains

Atoms

Residues

Water

Ligands

Bonds (RMSD)

Length (Å) (# > 4σ)

Angles (°) (# > 4σ)

MolProbity score

Clash score

Ramachandran plot (%)

Outliers

Allowed

Favored

Rama-2 (Ramachandran plot Z-score, RMSD)

whole (N = 4076)

helix (N = 512)

sheet (N = 1320)

loop (N = 2244)

Rotamer outliers (%)

CB outliers (%)

Peptide plane (%)

Cis proline/general

Twisted proline/general

CARAM outliers (%)

ADP (B-factors)

Iso/Aniso (#)

min/max/mean

Protein

Nucleotide

Ligand

Water

Occupancy

Mean

occ = 1 (%)

0 < occ < 1 (%)

occ > 1 (%)

12

34424 (Hydrogens: 0)

Protein: 4084 Nucleotide: 0

1464

MG: 8

0.004 (4)

0.026 (0)

2.13

6.30

0.00

2.55

97.45

-0.91 (0.13)

-0.44 (0.22)

-0.19 (0.13)

-0.85 (0.13)

7.67

0.00

8.1/0.3

0.0/0.0

2.09

34424/0

2.19/121.78/41.55

39.73/60.25/48.26

9.60/85.64/41.96

0.99

98.09

0.93

0.00

Data

Box

Lengths (Å)

Angles (°)

Supplied Resolution (Å)

Resolution Estimates (Å)

d FSC (half maps; 0.143)

d 99 (full/half/half2)

d model

d FSC model (0/0.143/0.5)

Map min/max/mean

Model vs. Data

CC (mask)

CC (box)

CC (peaks)

CC (volume)

Mean CC for ligands

Masked

Unmasked

3.1/---/---

3.2

2.9/2.9/3.1

-1.66/3.00/0.02

0.85

0.71

0.69

0.83

0.66